

## Raman Spectra of DNA in Aqueous Solution

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The infrared absorption spectra of DNA and polynucleotide have been studied by several workers,<sup>1-3</sup> but little work has been reported on the Raman spectra of an aqueous solution of DNA. In this note the author will report on the Raman spectra of an aqueous solution of DNA.

The calf-thymus DNA was purchased from the Sigma Chemical Co. The concentrations used were ca. 2 mg/25 ml tris buffer solution at pH=7.4. In order to cancel out the background due to the solvent molecules, tris buffer solutions were used as the reference substances. Before measurement the solutions were centrifuged at  $1 \times 10^4$  g for 60 min. All the measurements were made at room temperature. The excitation wavelengths were in the vicinity of 260 m $\mu$  and of 400 m $\mu$ . A slightly-modified Hitachi photoelectric spectrofluorometer was used for both the selection of the excitation ray and the analysis of the scattering spectrum. No fluorescence was observed in these measurements. To obtain the Raman spectra, the correction for Rayleigh scattering was made by the use of Eq. (1)<sup>4,5</sup>:

$$I_{\theta=\pi/2}(\lambda) = \left\{ \frac{2\pi^2 n_0^2}{N r^2 \lambda^4} \right\} \left( \frac{dn}{dc} \right)^2 G M^2 / z^2 \{ z - 1 + e^{-z} \} I_0(\lambda) \quad (1)$$

$$z = 2/3 \left\{ \frac{2\pi R n_0}{\lambda} \right\}^2 \sin^2 \left( \frac{\pi}{4} \right)$$

where  $I_{\theta=\pi/2}(\lambda)$  is the relative intensity of the Rayleigh scattering in the direction of  $\theta=\pi/2$ ;  $I_0(\lambda)$  the relative intensity of the incident ray;  $\lambda$ , the wavelength;  $r$ , the constant determined from the instrument;  $N$ , the Avogadro number;  $c$ , the concentration of the sample solution (g/ml);  $M$ , the molecular weight;  $R$ , the end-to-end distance of the random-coiling DNA,<sup>6</sup> and  $n_0$  and  $n$ , the refractive indices of the solvent and of the solution, respectively.

The corrected scattering spectrum of DNA

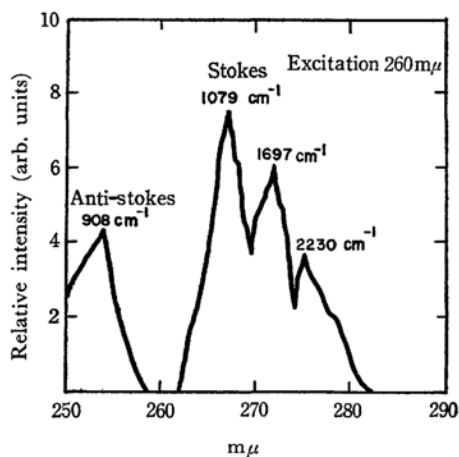


Fig. 1. Raman spectrum of DNA solution.

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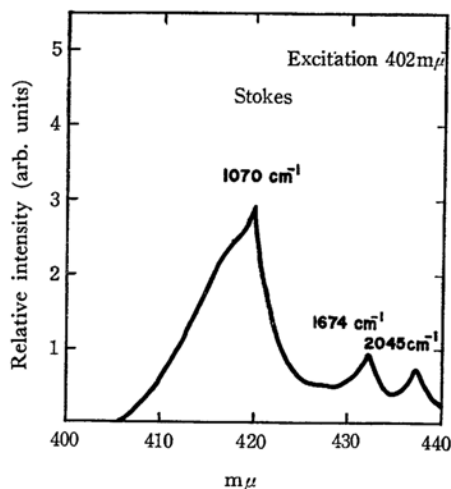


Fig. 2. Raman spectrum of DNA solution.

excited at 260  $m\mu$  is illustrated in Fig. 1, in which four frequencies,  $\tilde{\nu}_1 \approx 908 \text{ cm}^{-1}$ ,  $\tilde{\nu}_2 \approx 1079 \text{ cm}^{-1}$ ,  $\tilde{\nu}_3 \approx 1697 \text{ cm}^{-1}$  and  $\tilde{\nu}_4 \approx 2230 \text{ cm}^{-1}$ , are characteristic. In Fig. 2 a similar spectrum excited at 402  $m\mu$  is displayed. From the figure it is obvious that three frequencies,  $\tilde{\nu}_2 \approx 1070 \text{ cm}^{-1}$ ,  $\tilde{\nu}_3 \approx 1674 \text{ cm}^{-1}$  and  $\tilde{\nu}_4 \approx 2045 \text{ cm}^{-1}$ , are very similar to those of Fig. 1. These facts lead us to the conclusion that the corrected scattering spectra presented here are the Raman spectra of DNA in solution. That the anti-Stokes line,  $\tilde{\nu}_1 \approx 908 \text{ cm}^{-1}$ , was not observed in the spectra may be attributed to the error arising from the correction of Rayleigh scattering, because the spectral intensity of the scattering ray deviates from the Gaussian distribu-

TABLE I. RAMAN FREQUENCIES OF DNA IN AN AQUEOUS SOLUTION

Frequency, $\text{cm}^{-1}$	Assignment
$\tilde{\nu}_1$ 908	Sym. [OPO] stretching
$\tilde{\nu}_2$ 1070—1079	Sym. $[\text{PO}_2]^{-1}$ stretching
$\tilde{\nu}_3$ 1674—1697	C=C, C=N stretching in the purine ring
$\tilde{\nu}_4$ 2045—2230	?

tion. On the basis of these results, the Raman frequencies of DNA in a tris buffer solution were estimated to be as shown in Table I. The assignment of the frequencies listed in Table I based on the data of the infrared spectra of the purine base,<sup>7</sup> poly-A, DNA,<sup>8,9</sup> and on the Raman spectra of dimethyl phosphate anions.<sup>10</sup>

The Raman frequencies listed in Table I correspond well to the infrared absorption maxima of poly-A or DNA.<sup>2</sup> On the contrary, the chart produced by the commercial Raman spectrophotometer did not give any significant frequencies for these solutions. Therefore, the measurement of the Raman scattering described here would be a very useful tool for the study of macromolecules in solution.

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